

## BACKGROUND



Pushed by the necessity to boost drug development by giving access to relevant models, legislative bodies are expanding the usage of Organs-on-Chips (OoCs) submissions. There are still some significant challenges that need to be overcome to fully enable this transition. The lack of trained technicians in microfluidic operations, the high exigence of the industry (high throughput screening and high reproducibility), and the integration with their current technologies and readouts constitute some of these roadblocks. Here we present the development of our automated cell culture platform, a technological response to these issues.

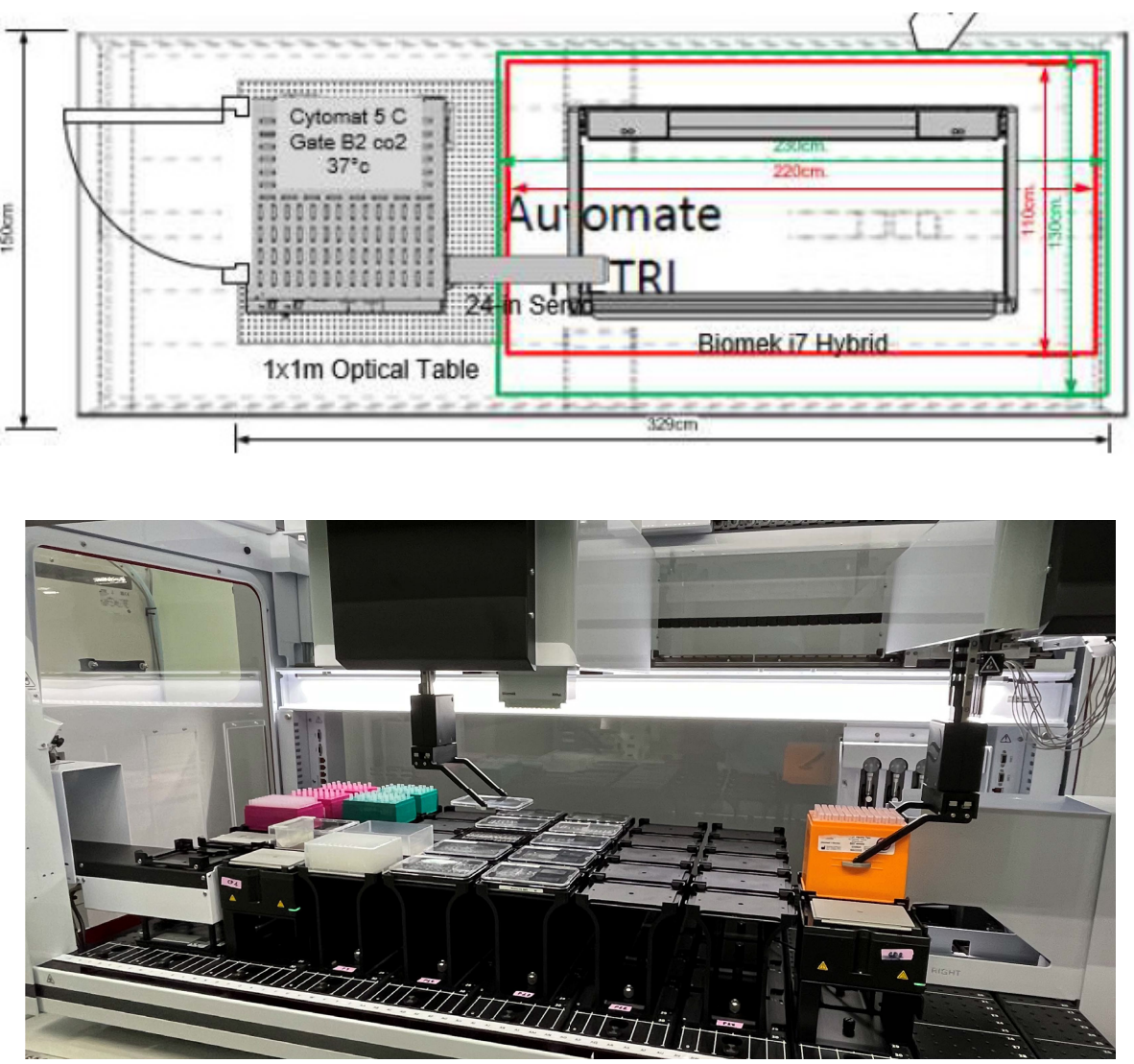
## MATERIAL

### AUTOMATED CELL CULTURE-PLATFORM.

To address this challenge, we present an automated cell culture-on-chip platform (Biomek 7, Beckman Coulter), and illustrate its impact on neuronal cells.



NETRI's microfluidic technology is compatible with automated cell culture.



### NETRI's microfluidic devices

- NeuroFluidics line
- In 96-well SBS format, NeoBento™

### Cells

- Rat Primary Hippocampal neurons

### Methods

- Cells were maintained in culture for more than three weeks
- Obtained manually versus using automated cell culture-platform

### Competitor

- Human laboratory technician (Manual)

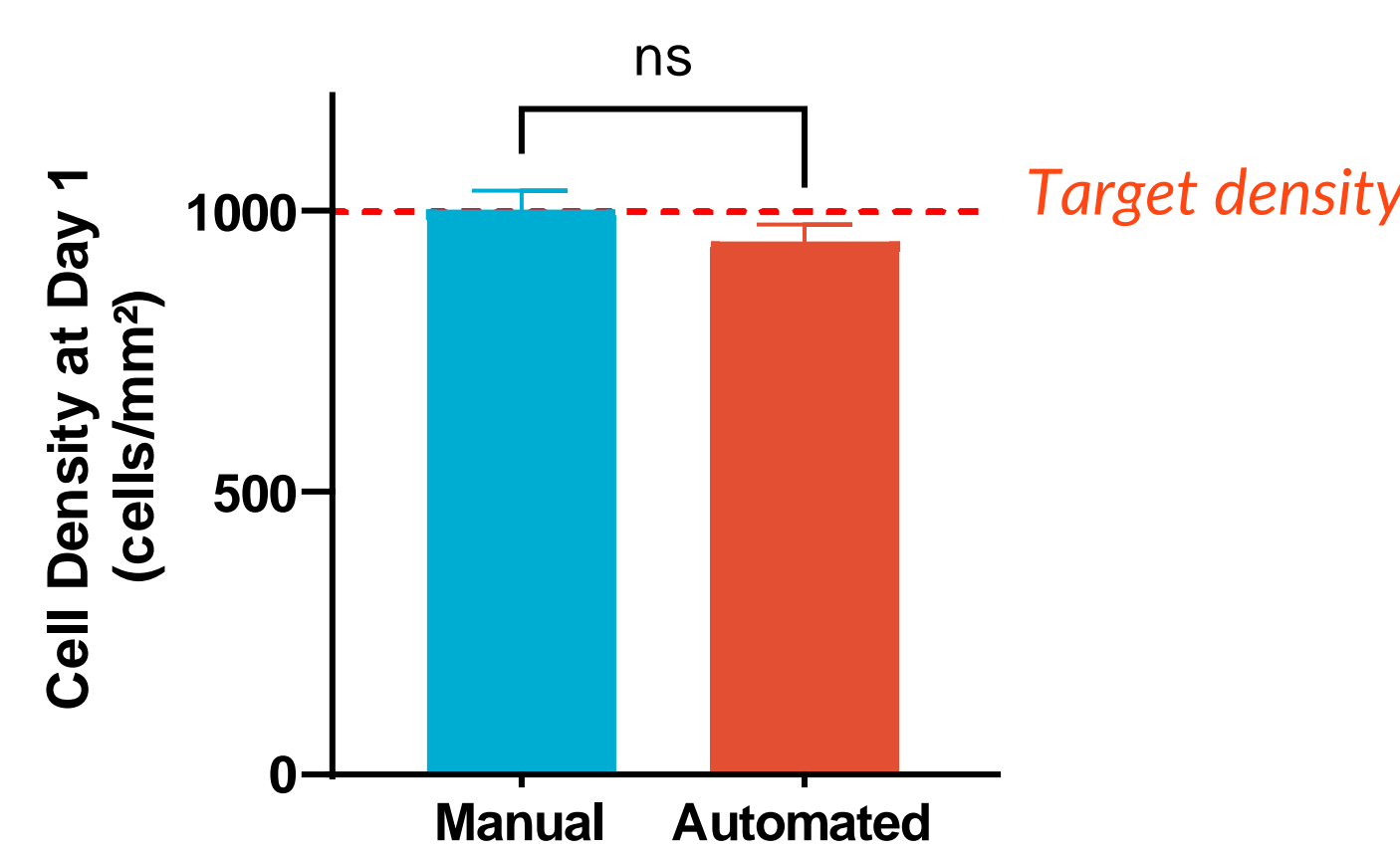


## RESULTS

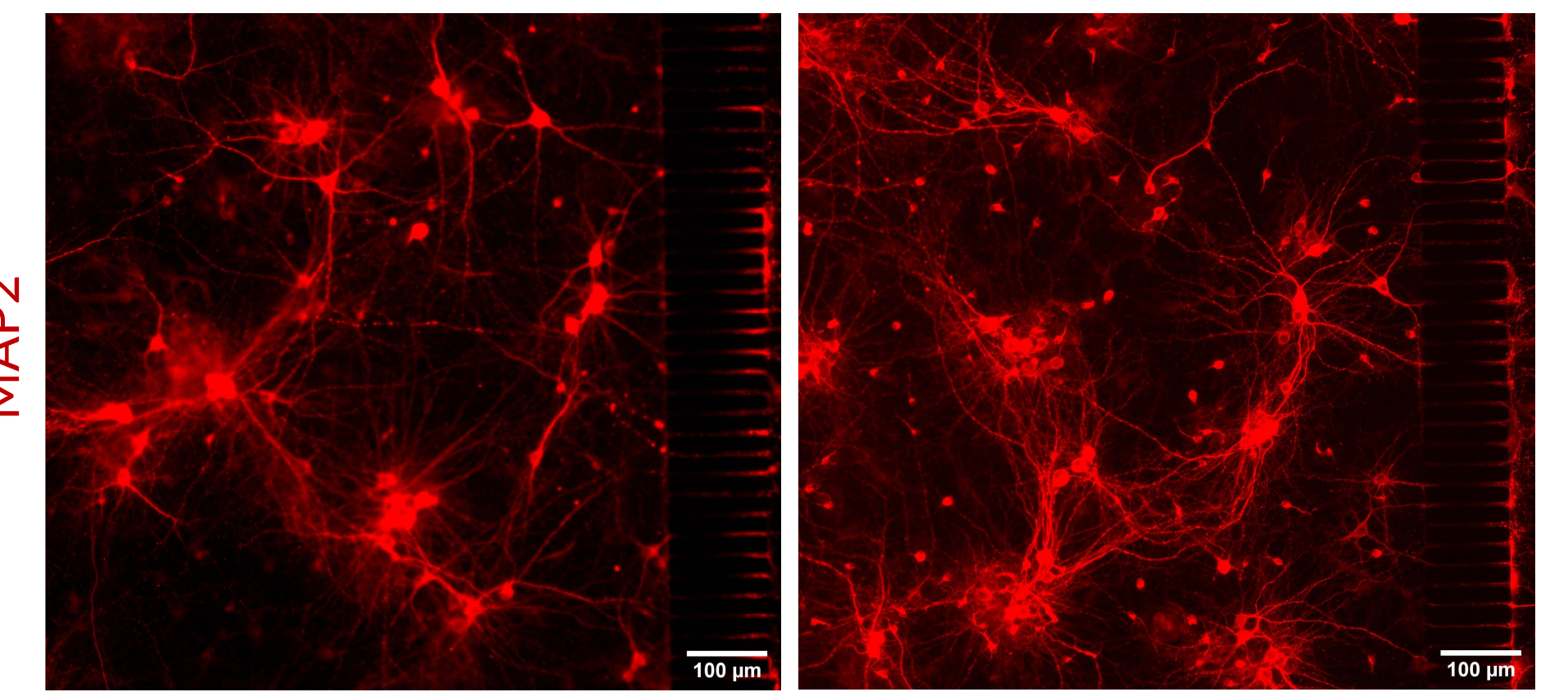
### COMPARABLE DATA BETWEEN MANUAL AND AUTOMATIC CULTURE

#### Biological relevance with automated cell culture-on-chip platform.

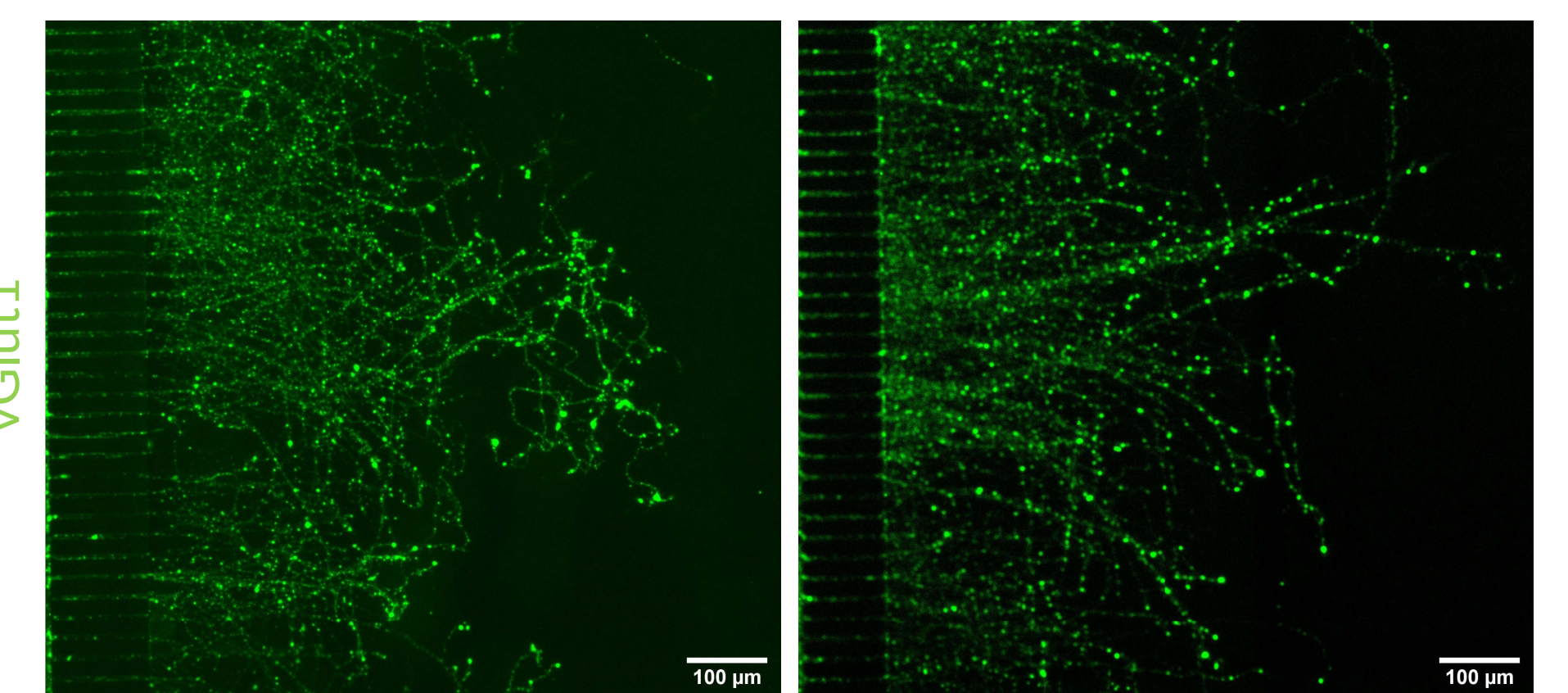
- Standard Operating Protocol enabling consistent cell seeding density
- Cell viability up to day 23
- Following axonal growth during culture
- Neuronal phenotypic expression of: vGlut1, MAP2,  $\beta$ III-Tubulin,



Quantification of cell density using automatic proprietary software (n=16, N=4 runs).



Channel 1  
MAP2

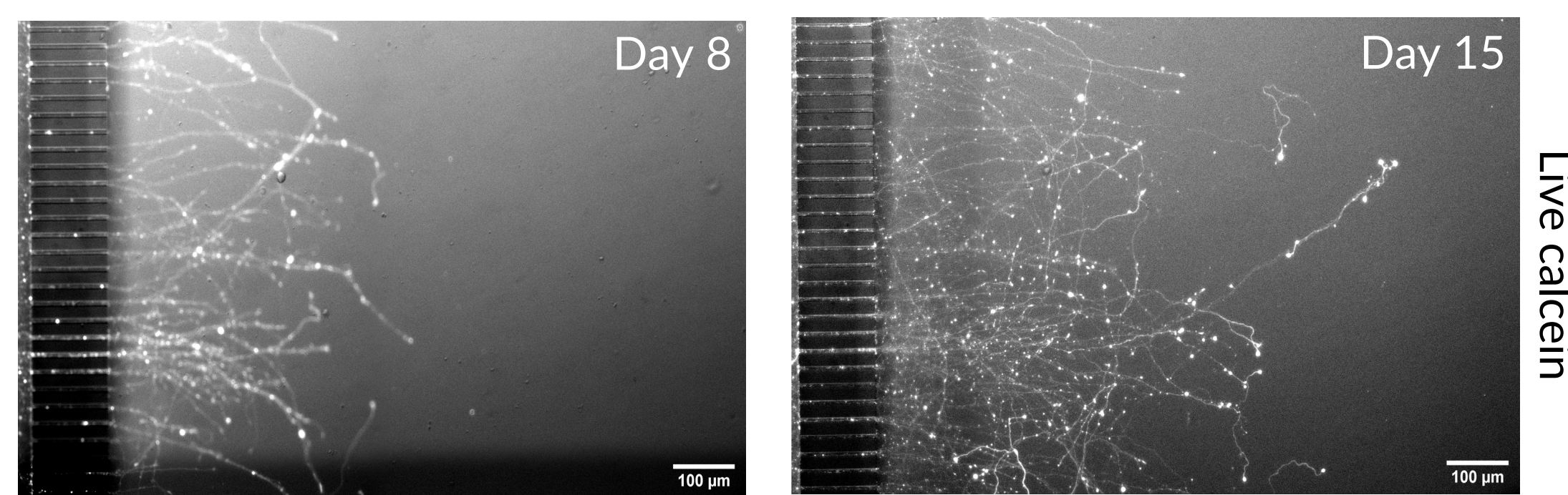


Channel 3  
vGlut1

Manual

Automated

Immunofluorescence pictures of primary hippocampal neurons cultured in microfluidic devices at Day 23 (n=3, N=4 runs).

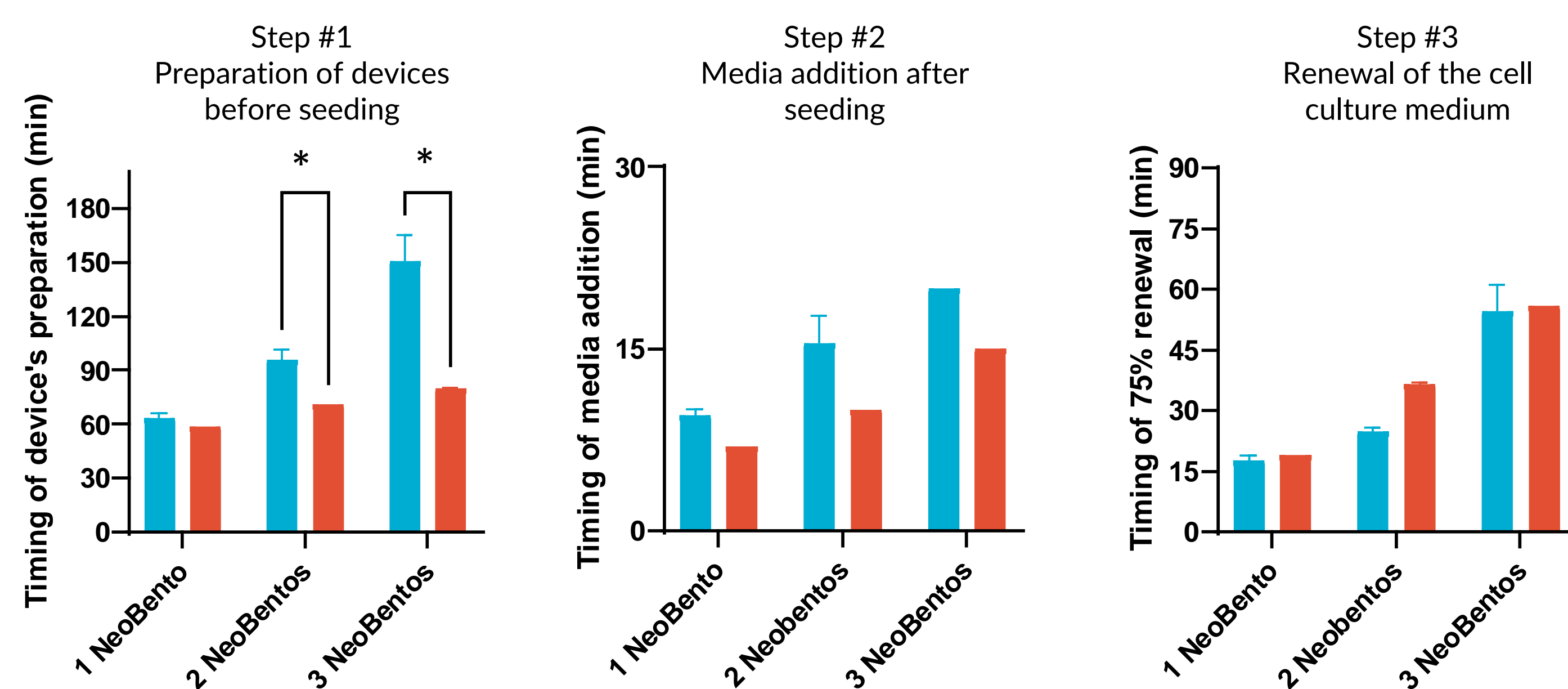


Dynamic neurite outgrowth of rat primary hippocampal neurons with Calcein live staining (0.1 µg/mL)

### HIGH THROUGHPUT STANDARDIZED NEURO-ORGAN-ON-CHIP.

#### Processing time and high throughput capacity thanks to automated cell culture platform:

- Time-saving at various stages
- Reproducible time at each stage of the cell culture protocol
- Improved reproducibility

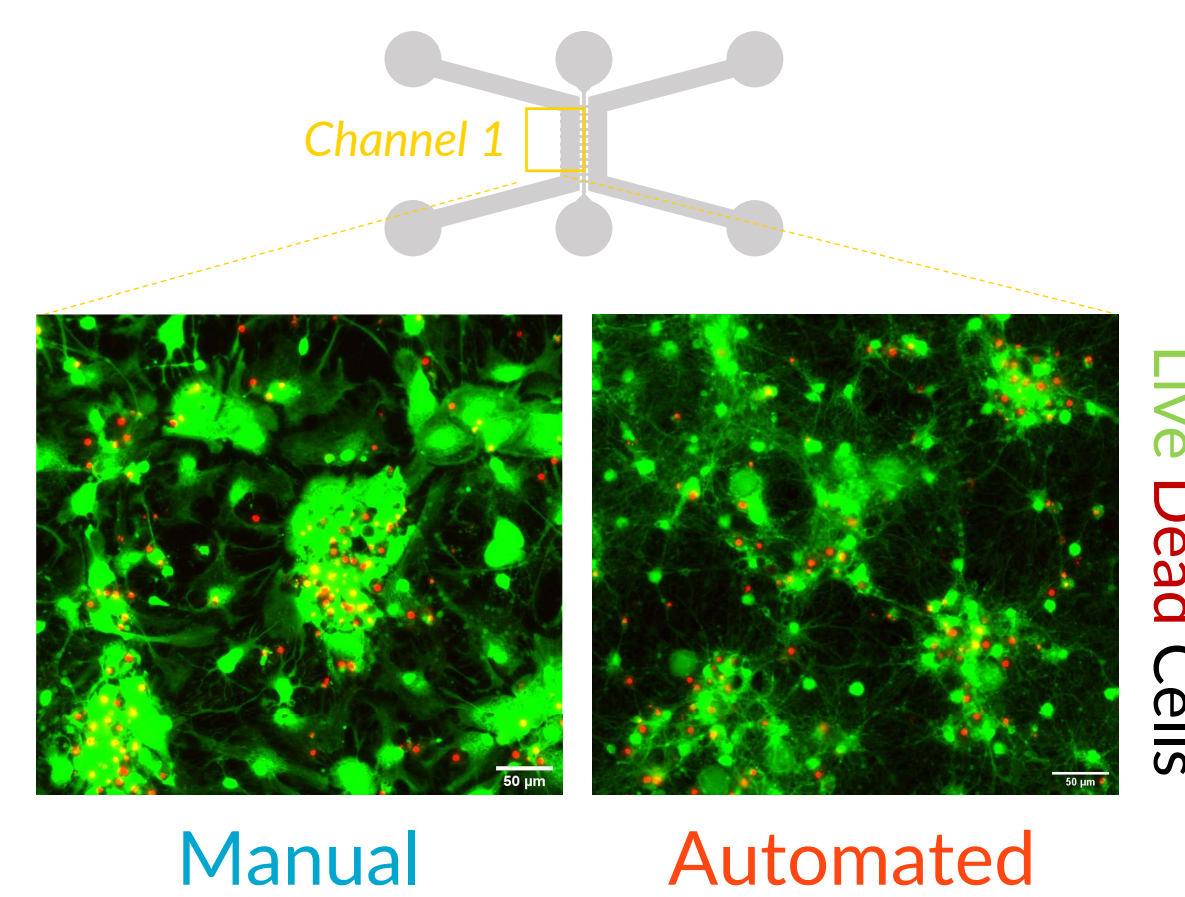


Automated  
Manual (n=3)

Timing of the experiment for three stages of the cell culture protocol (unpaired t-test, \*p-value <0.05).

#### Automated cell culture improves the viability of neuronal cells.

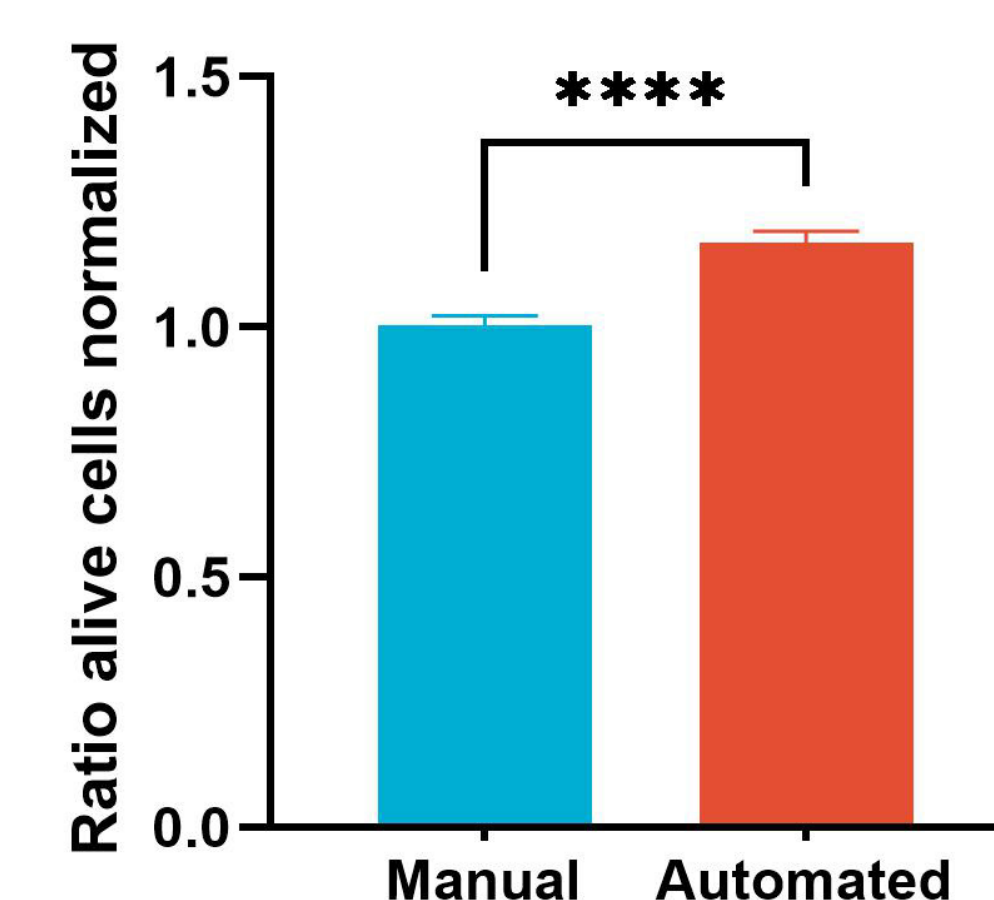
- Death/live quantification method
- Platform compatible with non-invasive analysis in the supernatant



Manual

Automated

Illustrative pictures of primary hippocampal neurons in channel 1 at day 23.



Quantification of alive cells at Day 23 using automatic proprietary software (n=7, N=4 runs, unpaired t-test, \*\*\*\*p-value <0.0001).

## CONCLUSION & PERSPECTIVES

### Approval of our tool and automated cell culture method

- Biological relevance, repeatability, reproducibility of models, and user experience efficiency
- Quality control to ensure model conformity
- High throughput capacity

### A production line for tomorrow's products to

- Access to pre-seeded human organs-on-Chip
- Increase the number of readouts (particularly for real-time monitoring)
- Use human cells derived from iPSCs

### Continuous improvement

- Optimization of automated protocols to move away from humans (i.e., for cell culture medium renewal step #3)
- Development of online quality control

BOOTH #926

